

Inoculation of Wheat With *Azospirillum* spp.: A Comparison Between Foliar and In-furrow Applications

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Received: September 19, 2019

Accepted: November 3, 2019

Online Published: December 15, 2019

doi:10.5539/jas.v12n1p194

URL: <https://doi.org/10.5539/jas.v12n1p194>

The research is financed by the CAPES (Brazil).

Abstract

The present work aimed to evaluate the agronomic performance of wheat grown under no-tillage system and submitted to different doses and methods of inoculation with *Azospirillum* spp. Eight treatments were tested: 1) control; 2) half N dose; 3) full dose of N; 4) half N dose + standard seed inoculation (200 mL ha⁻¹); 5) half N dose + in-furrow inoculation (200 mL ha⁻¹); 6) half N dose + in furrow inoculation (300 mL ha⁻¹); 7) half N dose + inoculation by foliar spraying (200 mL ha⁻¹) and 8) half N dose + inoculation by foliar spraying (300 mL ha⁻¹). The following parameters were evaluated: number of tillers per plant, number of grains per spike, shoot dry biomass, thousand seed mass, hectoliter weight, nitrogen content in grains and in shoot dry matter as well as crop yield. Our results showed that the supply of the half dose of mineral N associated to foliar inoculation with *Azospirillum* at the dose of 300 mL ha⁻¹ provided positive results on wheat yield, confirming the bacterial ability to fix N. However, only the full mineral N fertilization stood out as the best N fertilization management.

Keywords: *Triticum aestivum*, fertilization, nitrogen fixation, yield

1. Introduction

Over the last decade, Brazil has imported approximately half of its domestic wheat (*Triticum aestivum*) demand (CONAB, 2019a). To improve crop cultivation competitiveness against other winter crops such as corn, a more cost-effective fertilization is pointed out as a crop management obstacle to improve wheat profitability, specially regarding nitrogen (N), the country's most yield-limiting nutrient (Hungria et al., 2005) whose formulation relies on the importation of raw materials (CONAB, 2019b).

In soil, less than half of the N fertilizer applied to fields turns out to be available to plants, as N undergoes several loss processes, such as mineralization, immobilization, leaching and volatilization (Deuner et al., 2008). However, over the last 40 years, a wide array of investigations have confirmed the potential of the free-living bacteria *Azospirillum* spp. as fertilizer that, besides N fixation (Dobereiner & Day, 1976; Hungria et al., 2010) is also able to promote plant growth through the production of phytohormones (Dobbelaer et al., 1999; Fukami et al., 2018). In fact, wheat and corn management based on partial mineral N fertilization coupled with *Azospirillum* inoculation has shown to be able to meet the entire N requirement of these crops (Hungria et al., 2010; Cunha & Caierão, 2014; Marks et al., 2015).

As a phytosanitary strategy, oftentimes wheat seeds are treated prior sowing with plant protection products that, in the case of soybean, have shown to be harmful to the survival of the N-fixer bacteria *Bradyrhizobium* spp. (Campo et al., 2009). Although it has been poorly investigated; this toxicity is also likely to be found in the case of *Azospirillum* spp. In this context, besides the standard seed and in-furrow applications, as free-living bacteria *Azospirillum* spp. can theoretically further be explored through foliar sprayings (Fukami et al., 2018). This has

already been investigated by Fukami et al. (2016), who reported that both foliar and the in-furrow bacterial applications were as effective as standard seed inoculation to provide wheat with N.

In the Brazilian Paraná state, the main country's region of wheat production (CONAB, 2019a), this crop is cultivated under no-tillage system in succession to soybean harvesting, conditions in which almost no work with *Azospirillum* has been carried out. Therefore, there is a need for further studies aiming at investigating inoculation methods that, compared to seed inoculation, minimize or avoid the direct contact between plant protection products and bacterial inoculum. In this sense, this study particularly addressed the efficiency of foliar and in-furrow inoculation of wheat grown in a field rich in soybean residues that, as organic matter, act as N source, nutrient whose availability may impair the bacterial N fixation.

2. Material and Methods

The experiment was carried out at the institution experimental station (Lat. 23°02' S, Long. 52°04' W, and Alt. 509 m) of the *Universidade Estadual de Maringá* (UEM), Maringá-PR, southern Brazil. The soil type of the area is classified as dystroferic Red Argisol (Embrapa, 2018) and had previously been cultivated with soybean. The results of the chemical analysis in the 0-20 cm layer prior installation was: pH (CaCl₂) = 4.88, P (Mehlich-1 extraction) = 5.17 mg dm⁻³, H⁺ + Al³⁺ = 4.43 cmolc dm⁻³, Al³⁺ = 0.0 cmolc dm⁻³, K⁺ = 0.13 cmolc dm⁻³, Ca²⁺ = 2.75 cmolc dm⁻³, Mg²⁺ = 1.73 cmolc dm⁻³, CTC(T) = 9.04 cmolc dm⁻³ and BS = 51%. The soil texture of the same layer was as follow: 52% of sand, 3% of silt and 45% of clay.

The area is located under the Köppen a Cfa climate (humid subtropical) according to IAPAR (2018). The data from rainfall and maximum and minimum temperatures recorded by the in-site meteorological station over the crop cycle are resumed in the Figure 1.

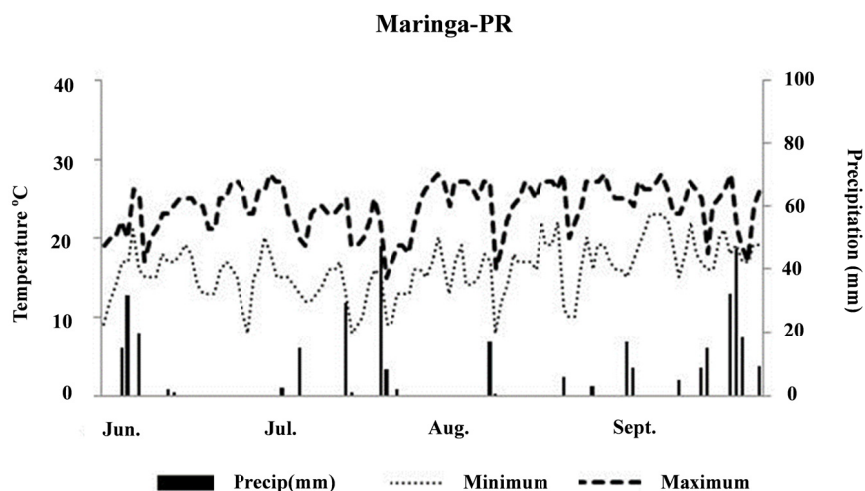


Figure 1. Summary of daily climatic data of rainfall and maximum and minimum temperatures over crop cycle (from June to September 2014). UEM's experimental station, Maringá-PR, Brazil

Adopting the no-tillage system, wheat cultivar CD 116 was sowed at the density of 420 viable seeds m⁻². Based on soil analysis, the crop fertilization and phytosanitary management were performed according to Cunha and Caierão (2014). The treatments consisted of the combination between two doses of inoculant and two methods of application as schemed in Table 1. Three other treatments were performed as comparison: the first was conducted without either N fertilizer or inoculant, the second consisted of half of the recommended N, whereas the third received the entire N input. Splitting one-third at sowing and the remaining at crop tillering, urea (46% of N) was used as mineral fertilizer, while a liquid formulation containing the *A. brasilense* AbV5 and AbV6 strains (concentration of 2×10^8 CFU mL⁻¹) was adopted as inoculant.

Table 1. Scheme of the treatments summarizing N fertilizer rates, *A. brasilense* doses and methods of applications in wheat cv. CD116

| Treatment | Mineral N fertilization (kg ha ⁻¹) | Doses of inoculant (mL ha ⁻¹ /100 kg ⁻¹ seeds) | Methods of inoculation |
|-----------|--|--|-------------------------------|
| 1 | 0 | 0 | - |
| 2 | 55 | 0 | - |
| 3 | 110 | 0 | - |
| 4 | 55 | 200 | Seed application ¹ |
| 5 | 55 | 200 | In-furrow ² |
| 6 | 55 | 300 | In-furrow ² |
| 7 | 55 | 200 | Foliar spraying ³ |
| 8 | 55 | 300 | Foliar spraying ³ |

Note. ¹ Seed coating using 200 mL 100 kg⁻¹; ² Slurry volume of 50 L ha⁻¹; ³ Slurry volume of 150 L ha⁻¹.

The experiment was performed in a completely randomized block design with eight treatments and four replications. Each plot was composed by 10 rows of 5.0 m length, spaced 0.16 m apart. At harvest, two outer lines were removed, as well as 0.5 m from each row end. Only plants from the six central rows of each experimental unit were evaluated, totalizing a useful area of 3.84 m². The variables analyzed were as follows:

Shoot dry biomass: It was determined at the beginning of the flowering stage by randomly collecting 10 plants from the useful area of each plot. After dried in a forced air oven at 65 °C until constant weight, the biomasses were weighted and the results were expressed as the average of 10 plants.

Number of tillers per plant: 10 plants per plot were evaluated at the beginning of the boot stage.

Numbers of grains per spike: It was determined by collecting 10 spikes per plot at the crop complete maturation stage.

Crop yield: It was obtained from the grain mass harvested from the useful area of each plot. Yield was expressed in kg ha⁻¹ at 13% of moisture (Brasil, 2009).

Thousand seed mass: It was performed by weighing eight subsamples of 100 seeds for each replication according to Brasil (2009).

Hectoliter weight: It was determined by weighing a known volume (225 mL) of grains in a portable hectoliter device (Agrologic AI-101[®]). The results were expressed in kg hL⁻¹ (Brasil, 2009).

Nitrogen content in the shoot and grains: It was performed applying the Kjeldahl method (AOAC, 2010) to the samples collected to evaluate shoot dry biomass and yield, respectively.

Statistical analysis: All data were tested for residual homoscedasticity and normality and submitted to analysis of variance. When significant, means were compared by Duncan's test ($p \leq 0.05$) to identify pair of means and determine the significant differences between them (SAS, 2014).

3. Results and Discussion

The analysis of variance showed that N fertilization combined with *Azospirillum* spp. significantly altered wheat shoot dry biomass (SDB), N content in dry shoot (NCDS), N content in grains (NCG) as well as crop yield (CY). On the other hand, the number of tillers per plant (NTP), number of grains per spike (NGS), hectoliter weight (HW) and thousand seeds mass (TSM) were not significantly affected by the treatments described in Table 2.

Table 2. Mean values of number of tillers per plant (NTP), number of grains per spike (NGS), hectolitic weight (HW), shoot dry biomass (SDW), thousand seeds mass (TSM), N content in dry shoot (NCDS), N content in seeds (NCG) and crop yield (CY) of wheat cv. CD116, in response to different methods and doses of inoculant. Maringá-PR, Brazil

| Treatments | NTP | NGS | HW (kg hL ⁻¹) | SDW (g plant ⁻¹) |
|------------|---------------------|----------------------------------|------------------------------|------------------------------|
| 1 | 2.84 ^{ns} | 41.60 ^{ns} | 78.35 ^{ns} | 0.863 b |
| 2 | 2.95 | 42.60 | 78.60 | 0.933 b |
| 3 | 2.75 | 42.50 | 78.65 | 1.425 a |
| 4 | 2.65 | 42.50 | 78.33 | 0.923 b |
| 5 | 2.90 | 42.50 | 78.58 | 0.808 b |
| 6 | 2.80 | 42.50 | 78.41 | 1.163 ab |
| 7 | 2.62 | 42.50 | 78.40 | 0.905 b |
| 8 | 2.68 | 42.50 | 78.50 | 1.185 ab |
| CV (%) | 22.7 | 8.5 | 2.3 | 24.9 |
| Treatments | TSM (g) | NCDS (mg N plant ⁻¹) | NCG (kg N ha ⁻¹) | CY (kg ha ⁻¹) |
| 1 | 40.61 ^{ns} | 3.858 b | 49.93 d | 2411.31 d |
| 2 | 40.51 | 5.193 a | 57.82 c | 2481.01 cd |
| 3 | 40.48 | 5.338 a | 113.51 a | 3410.56 a |
| 4 | 40.19 | 5.598 a | 56.68 c | 2553.29 c |
| 5 | 40.39 | 6.818 a | 55.24 cd | 2477.81 cd |
| 6 | 40.47 | 6.118 a | 68.06 b | 2494.63 cd |
| 7 | 40.59 | 5.663 a | 56.59 cd | 2490.25 cd |
| 8 | 41.00 | 6.933 a | 70.00 b | 2665.63 b |
| CV (%) | 2.4 | 16.3 | 3.1 | 9.9 |

Note. ^{ns} = not significant; Means followed by the same letter in the column do not differ statistically according to Duncan's test ($p \leq 0.05$).

Regarding the number of tillers, Salantur et al. (2006) pointed out this yield component as the main responsible for increasing wheat productivity in fields with not-established population of *Azospirillum* spp. However, differently from these authors as well as to Fukami et al. (2016), in the present work any yield promotion can be attributed to tillering, as NTP data was not significant (Table 2). Nunes et al. (2015) and Ferreira et al. (2017) also did not reported any increase of NTP as result of inoculation and attributed the crop lack of response to *Azospirillum* spp. to the soil high N availability prior sowing.

In this work, SDW data obtained from the in-furrow (treatment 6) and foliar (treatment 8) applications at the dose of 300 mL ha⁻¹ provided similar values to treatment 3, in which full N mineral fertilization was applied (Table 2). However, it is important to point out that our SDW results were not conclusive, as, at the same time, treatments 6 and 8 showed comparable values to the others, including the control. Therefore, our SDW results reinforce those of Fukami et al. (2016) and Hungria et al. (2010), who also reported no increase in wheat shoot biomass triggered by inoculation. In fact, diazotrophic bacteria have shown to be more efficient at stimulating shoot growth in soils poor in N (Salantur et al., 2006), since a high availability of this element can impair the activity of nitrogenase, the enzyme responsible for reducing atmospheric N into biologically available compounds (Fukami et al., 2018). In this sense, it is likely to presume that in our work the remaining or native bacterial population coupled with soybean residues from precedent seasons may have constituted an available N source to plants, which may have neutralized or even inhibited the bacterial stimulation of SDW, NTP, NGS, HW and TSM.

Comparing standard seed-inoculation to the in-furrow and foliar ones, Fukami et al. (2016) concluded that the two latter methods were more promising to increase *Azospirillum* abundance and N-content in plant tissues. Nonetheless, our NCDS results did not corroborate their outcome, as herein any dose or method of inoculation affected leaves N content (Table 2). Other than that, our results support Nunes et al. (2015), since NCDS clearly responded to N fertilization. Plant-bacteria symbiosis is a species-specific process (Droque et al., 2014) that, in the case of wheat inoculation with *Azospirillum*, has further shown to be genotype-dependent. Therefore, besides the soil N availability provided by the organic matter of soybean, it cannot be discarded the hypothesis that varietal differences could additionally have contributed to the NCDS results found in this work (Kaiz et al., 2016).

Another important aspect related to N content in wheat is the protein concentration of grains, since this is a key technological factor in flour quality (Cazetta et al., 2008). In this work, full supply of mineral N (treatment 3) presented the highest value of NCG (Table 2), while intermediate data were observed under the inoculant dose of 300 mL ha⁻¹ in both in-furrow and foliar applications (treatments 6 and 8, respectively). Such similar performance was also observed concerning the productivity, as only the treatment based on entire mineral fertilization provided the highest CY value (Table 2).

Unlike what we observed in this work, Hungria et al. (2010), Fukami et al. (2016), and Marks et al. (2015) reported that N management based on mineral fertilization coupled with standard (seed) or foliar inoculations provided wheat yields equivalent to those obtained under the full N recommendation. However, it is worthwhile mentioning that differently from those mentioned authors, in this work the N fertilizer rate was 25% lower. Despite that, our results confirm the potential of foliar *Azospirillum* application to partially contribute with the input of N to plants.

4. Conclusion

The supply of the half dose of mineral N associated to foliar inoculation with *Azospirillum* at the dose of 300 mL ha⁻¹ provided positive results on wheat yield. However, the full mineral N fertilization stood out as the best N fertilization management.

Acknowledgements

To *Coordenação de Aperfeiçoamento de Pessoal de Nível Superior*—CAPES for the scholarships awarded to the first author as well as for providing the resources for research development.

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